



A pipeline for studying minor variants in complex genetic populations using long reads from high-throughput sequencing technologies

Ortega-Serrano, I.; Quer, J.; Rodriguez-Frias F.; Sánchez-Pla, A.



Outline

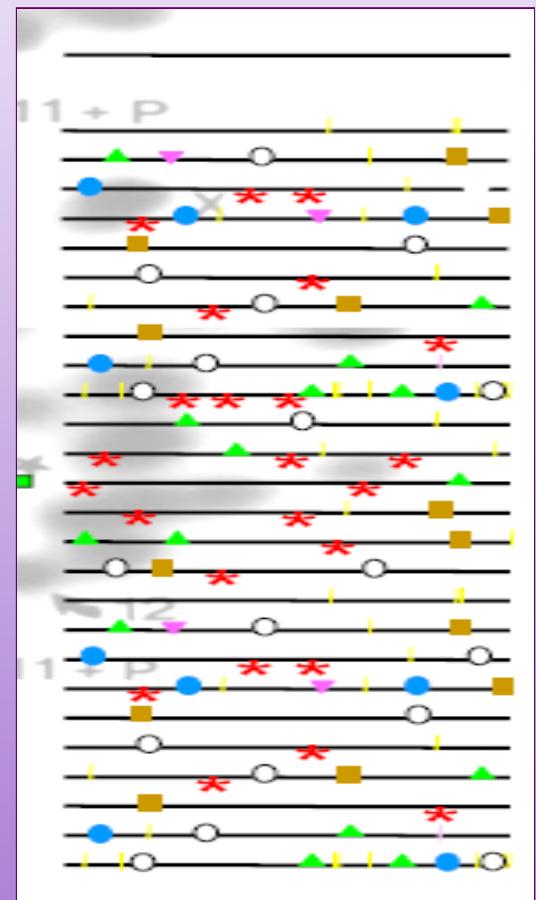
- Motivation and experimental goals
 - HCV, HBV viruses
- Why Next Generation Sequencing (NGS) technologies?
- Which NGS technology?
- Pipeline to process reads
 - Non-specific filter to remove low-quality reads
 - Statistical modelling of erroneous variants
- Results

Motivation and experimental goals

- Hepatitis C and Hepatitis B viruses circulate as quasispecies

In an infected patient:

- (1) The population of viruses presents high rates of mutation and replication. It is a complex mixing of different mutants.
- (2) These mutants (*variants*) are related amongst them
- (3) They are subjects to competition and natural selection.



Motivation and experimental goals

- › Goals of the study:
 - › Detection and quantification of **mutations** or **combination of mutations** that could confer resistance to viral inhibitors in samples from chronically infected patients.
 - › HCV: mutations in NS3 protease
 - › HBV: mutations in polymerase

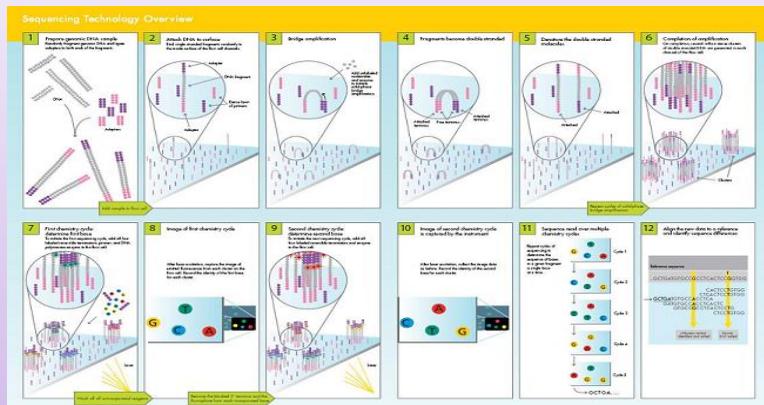
Why NGS technologies?

- Minor variants often play an important role in the development of resistance to antiviral treatments in patients, even if they are present in a very low percentage in the population.
 - Minor variants may not be detected by classical sequencing methods
 - You obtain hundreds of sequences with much effort and high cost
 - NGS tools allow to detect minor variants efficiently
 - You obtain thousands of sequences with relatively low cost

Which NGS technology?

- Solexa-Illumina GA/AB SOLiD/....

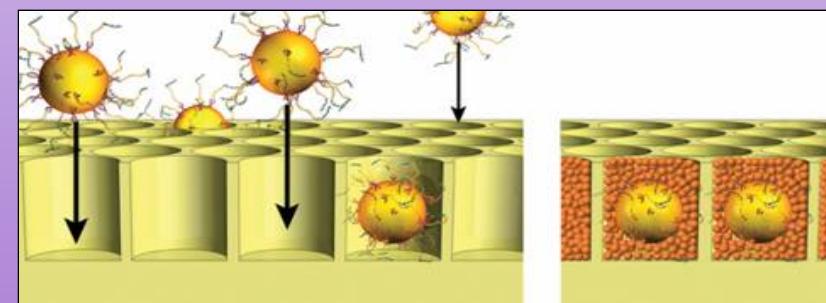
- 35-100 Short reads, useful for the study of SNPs or short regions



- 454-Roche GS FLX platform:

- With longer reads (250-400) a “wider picture” is obtained
- This is an advantage for the study of combination of mutations in a same sequence
- Artifactual indels in homopolymers:

```
...CGGCCGGGACAAAAACCAGGTGGA...
...CGGCCGGGACAAACCAGGTGGA....
...CGGCCGGACAAAAAACCAGGTGGA....
...CGGCCGGGACAAAAAACCAGGTGGA...
```



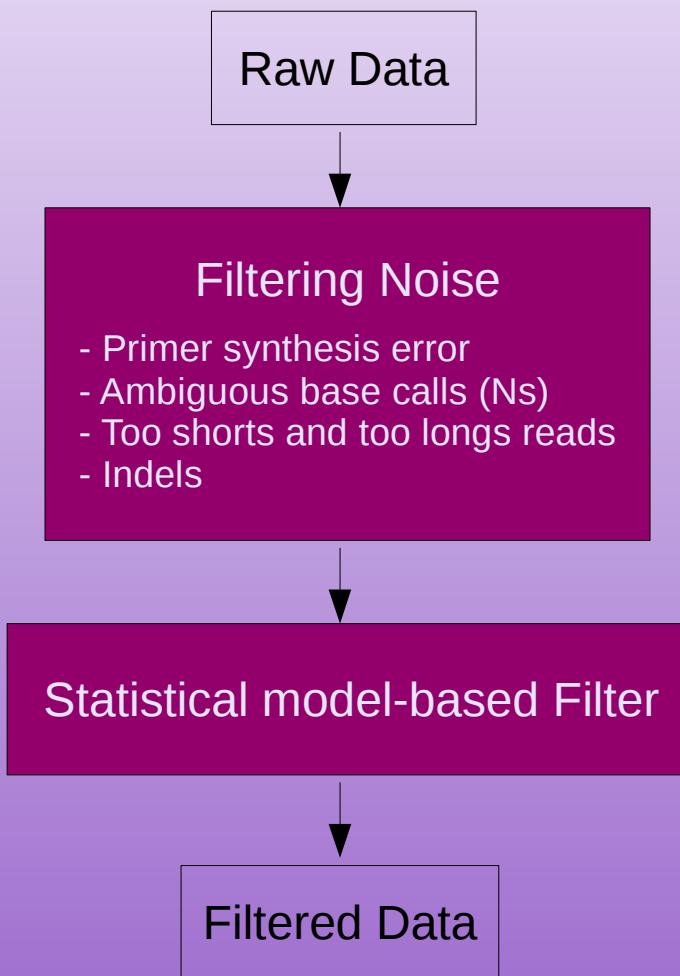
Nucleotide reference sequence of HBV polymerase:

5'

ATGTTTCCCTCATGTTGCTGTAC.AAA.ACC.TAC.GGA.TGG.AAA.TTG.CAC.CTG.TAT.TCC.CAT.CCC.ATC.GTC.CTG.GGC.TTT.CGC.AAA.
ATA.CCT.ATG.GGA.GTG.GGC.CTC.AGT.CCG.TTT.CTC.TTG.GCT.CAG.TTT.ACT.AGT.GCC.ATT.TGT.TCA.GTG.GTT.CGT.AGG.GCT.
TTC.CCC.CAC.TGT.TTG.GCT.TTC.AGC.TAT.ATG.GAT.GAT.GTG.GTA.TTGGGGGCCAAGTCTGTACAG 3'

Pipeline to process reads

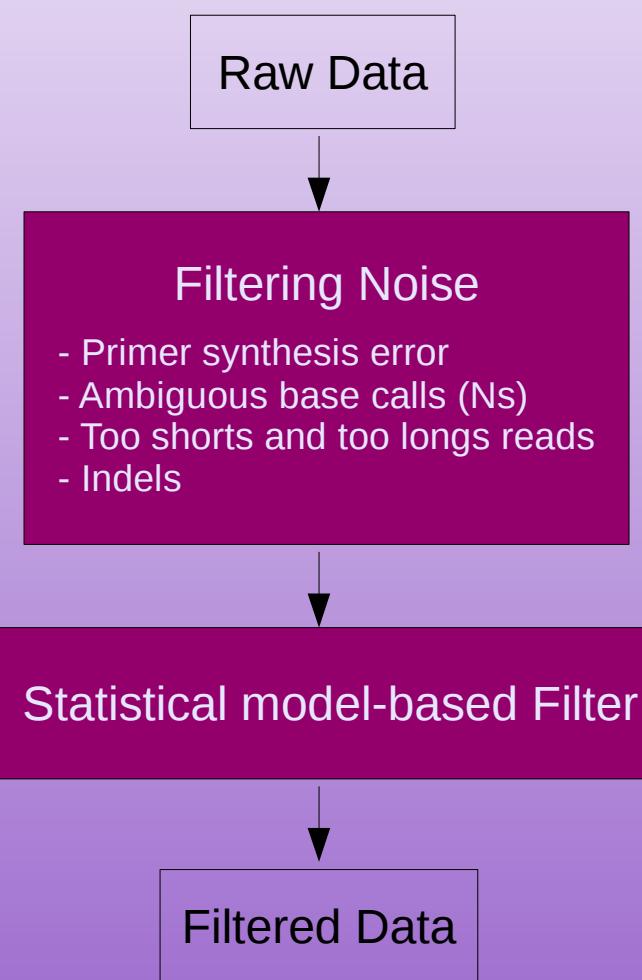
- Goal: to obtain accurate estimations of proportions of the variants.



- Two main steps:
 - Filtering low-quality reads more accurate estimations will be obtained.
 - Statistical model for the remaining error: modelling the errors that still remain in our reads.

Assessing the pipeline performance

- To check the performance of the pipeline:
 - Pyrosequencing 3 independent PCR products from a clon
 - As we know the sequence of the clon any change in the reads is considered a process error.



Nucleotide reference sequence of the clon:

5'
GCTGGCCGCTCCCCAAGGTGCCGCTCACTGACACCCTGCACTTGCGGCTCTCGGACCTTACCTG
GTCACGAGGCACGCCGATGTCAATTCCCGTACGCCGGGGGTGATGGCAGGGGCAGCCTGCTTCGCC
CCGGCCCACATCTTACCTGAAAGGCTCTCGGGGGGCCACTGCTGTGCCCGCGGGACACGCCGTAG
GCATTTCAAGAGCCCGGGTGTGCACCCGTGGAGTGGCTAAAGCGGTGGACTTTATCCCGTAGAGGGC
CTAGAGACAACCATGAGGTCCCC**GGTGTTC**CGGACAATTCTCC 3'

5'
GCTGGCCGCTCCCCAAGGTGCCGCT**T**CTGACACCCTGCACTT**CCGGCTCCTCGGACCTTACCTG**
GTCACGAGGCACGCCGATGTCAATTCCCGTACGCCGGGGGTGATGGCAGGGGCAGCCTGCTTCGCC
CCGGCCCACATCTTACCTGAAAGG**A**TCTCGGGGGGCCACTGCTGTGCCCGCGGGACACGCCGTAG
GCATTTCAAGAGCCCGGGTGTGCACCCGTGGAGTGGCTAAA**AGCGGTGGACTTTATCCCGTAGAGGG**
CCTAGAGACAACCATGAGGTCCCCGGTGTCTCGGACAATTCTCC 3'

Assessing the pipeline performance

- To check the performance of the pipeline:
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Raw Data

Filtering Noise

- Primer synthesis error
- Ambiguous base calls (Ns)
- Too shorts and too longs reads
- Indels

Statistical model-based Filter

Filtered Data

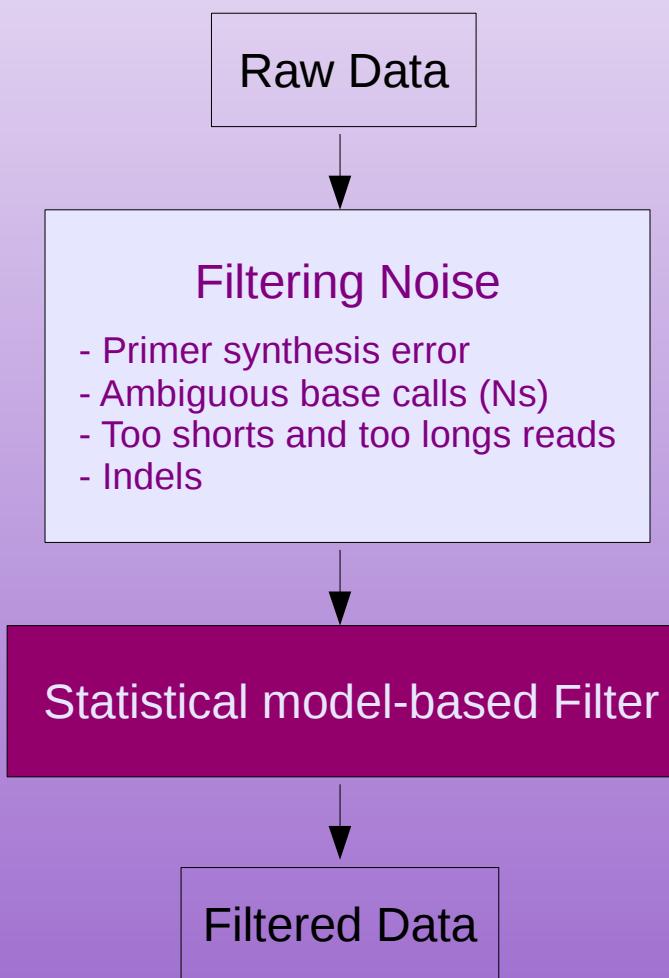
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GTCACGAGGCACGCCGATGTCAATTCCCGTACGCCGGGGGTGATGGCAGGGGCAGCCTGCTTCGCC
CCGGCCCACATCTTACCTGAAAGGCTCTCGGGGGGCCACTGCTGTGCCCGCAGGACACGCCGTAG
GCATTTCAAGAGCCGCGGTGTGCACCCGGTGGAGTGGCTAAAGCGGTGGACTTATCCCCGTAGAGGGC
CTAGAGACAACCAGAGGTCCCCGGTGTGGCTAAAGCGGTGGACTTATCCCCGTAGAGGGC

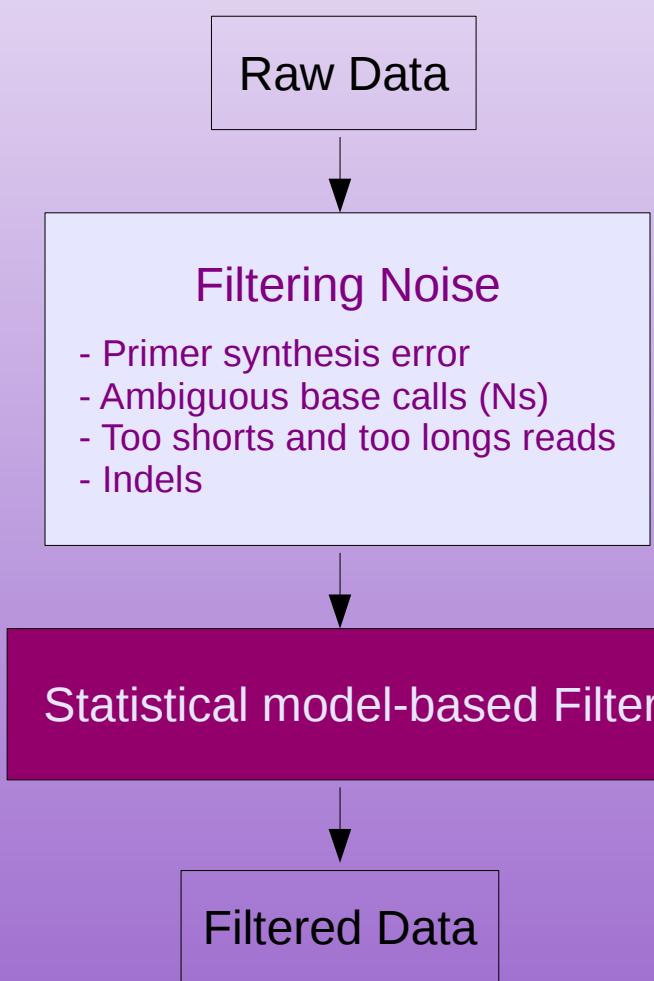
Process errors

Filtering noise

- Remove reads with more than 25% of primer synthesis errors



Filtering noise



- Remove reads with more than 25% of primer synthesis errors
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Filtering noise

Raw Data

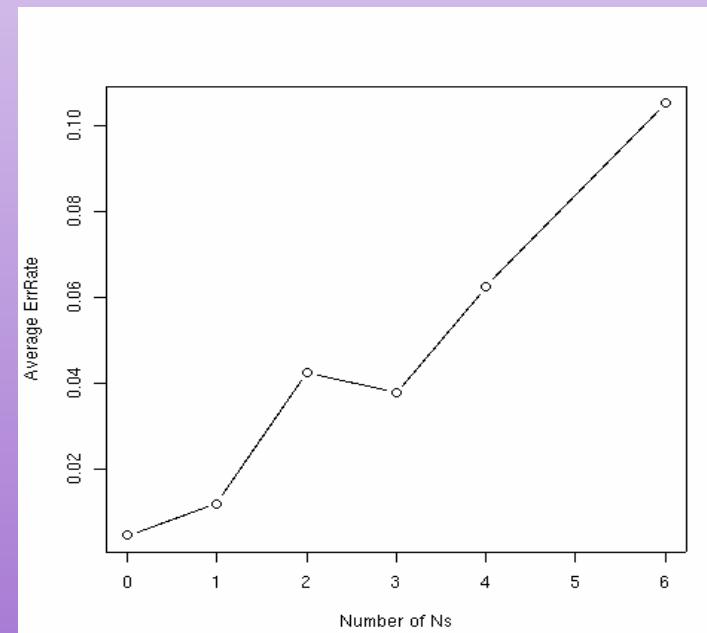
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Filtering Noise

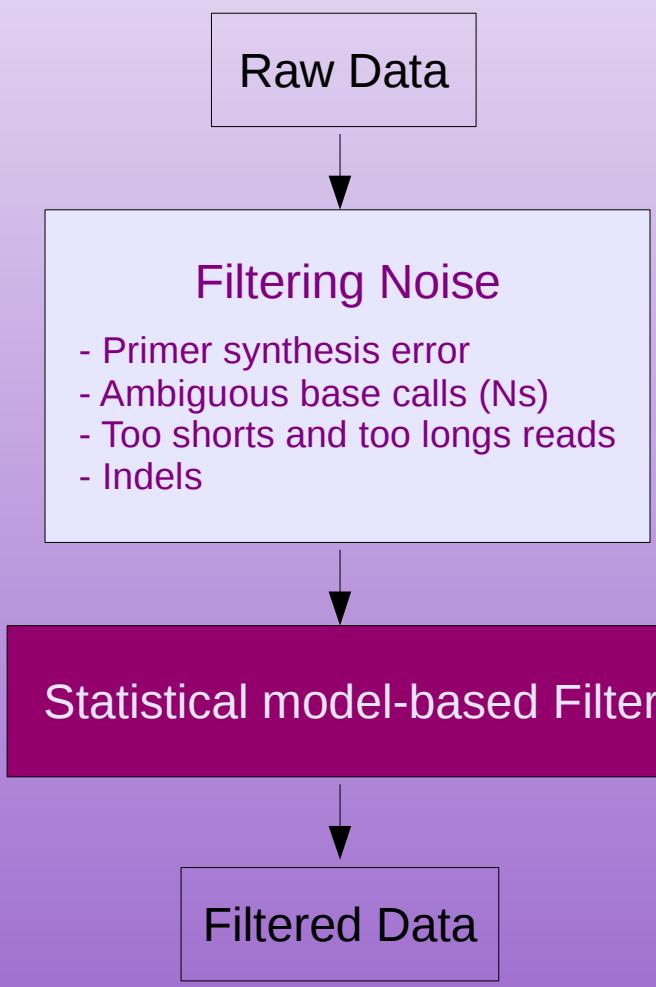
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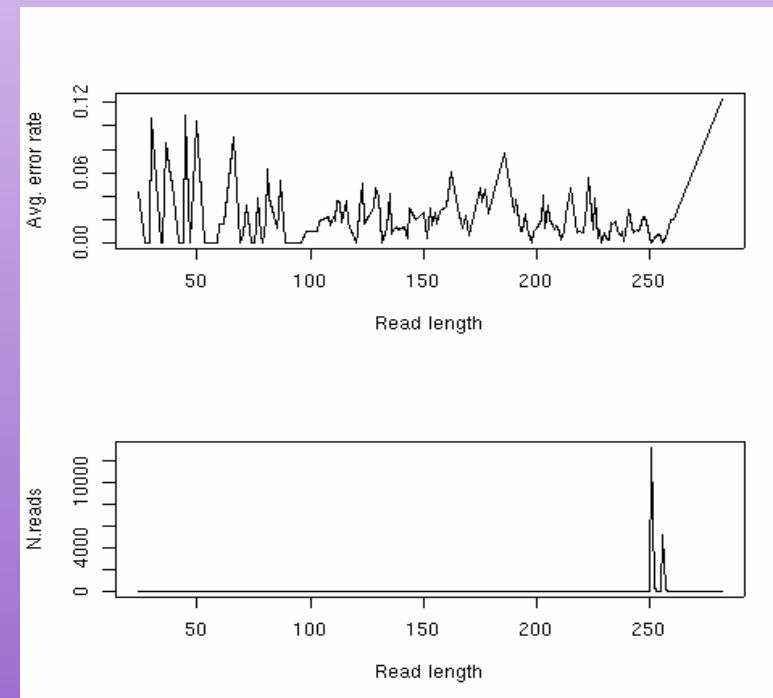
Filtered Data



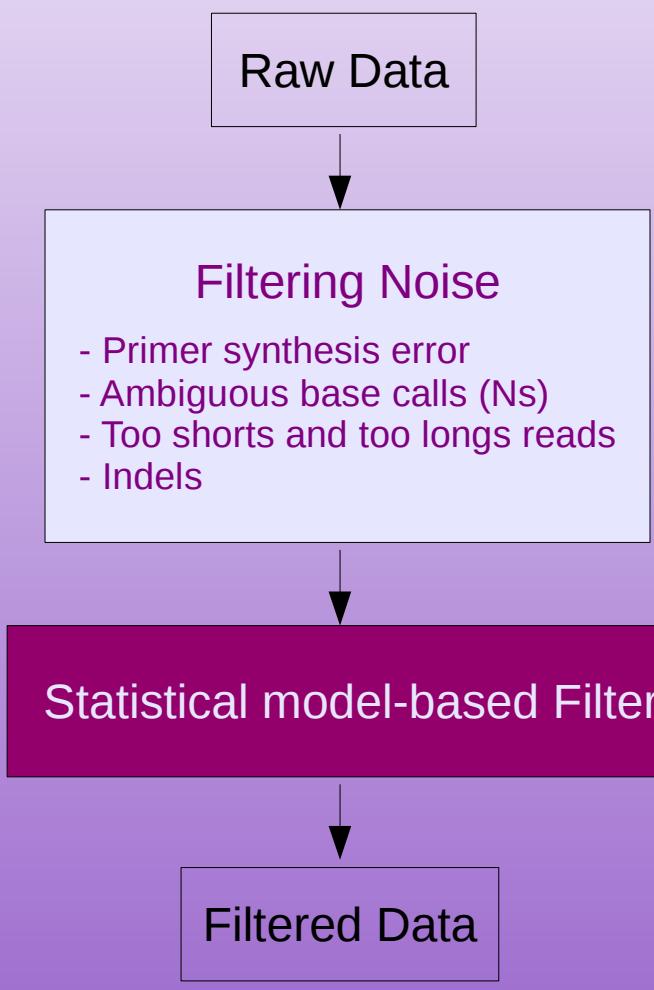
Filtering noise



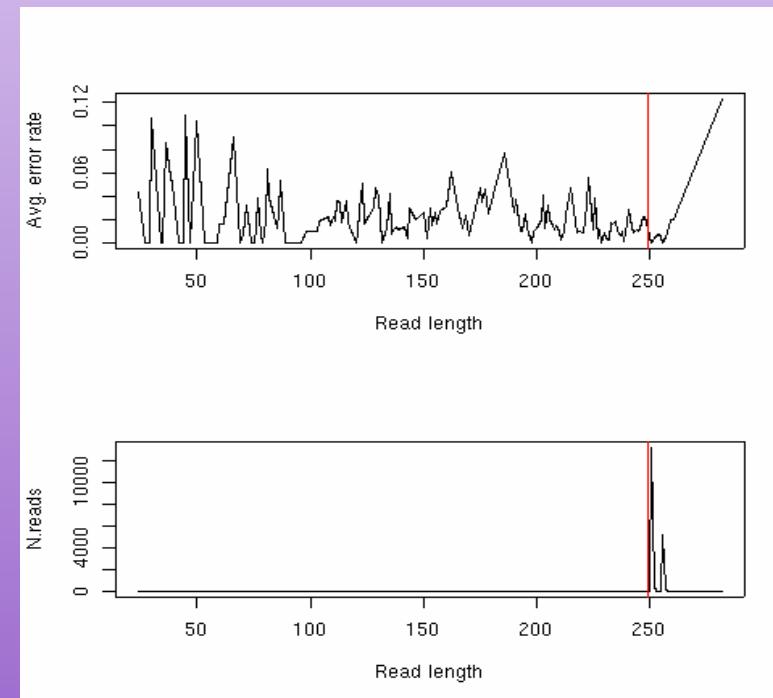
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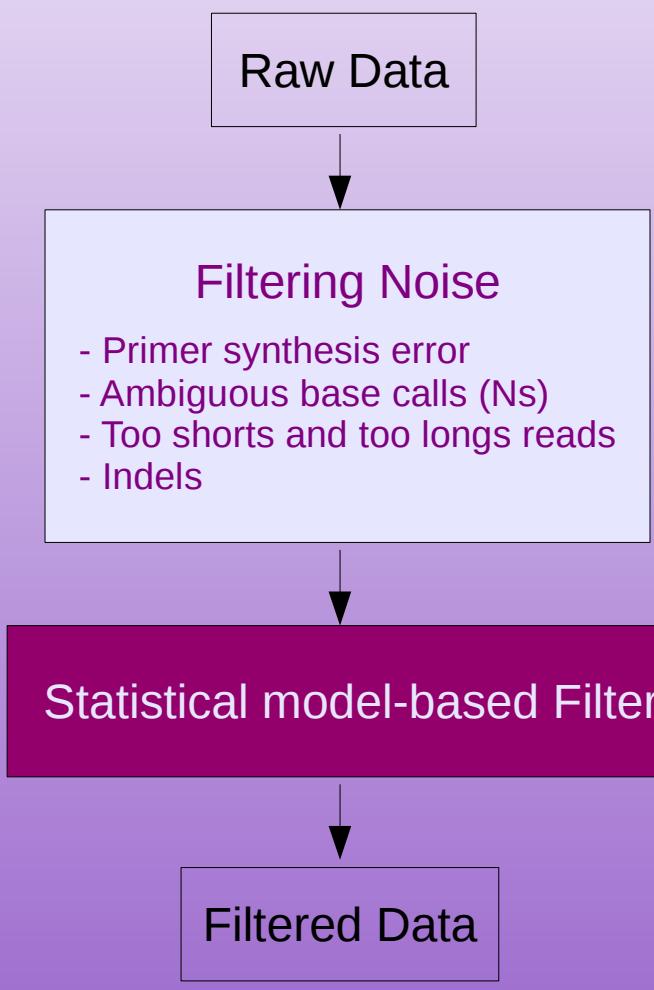
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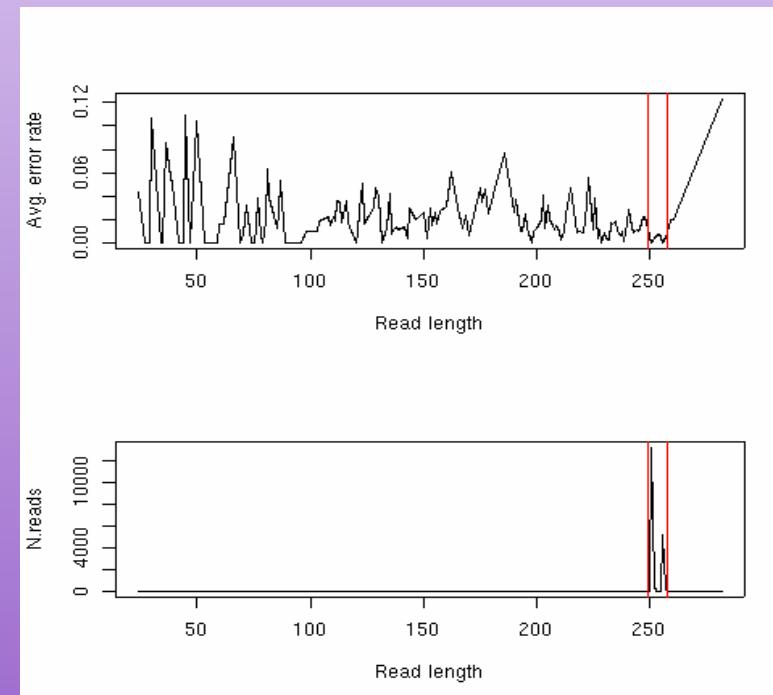
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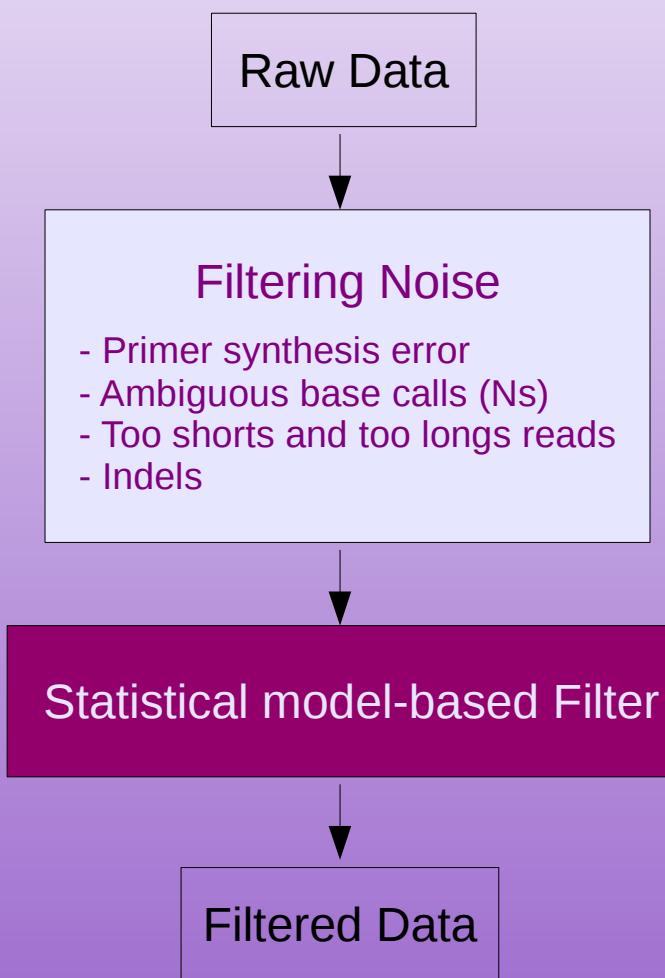
Filtering noise



- Remove reads with more than 25% of primer synthesis errors
- Remove reads with ambiguous base calls (N)
- Remove short reads and trim too long ones:



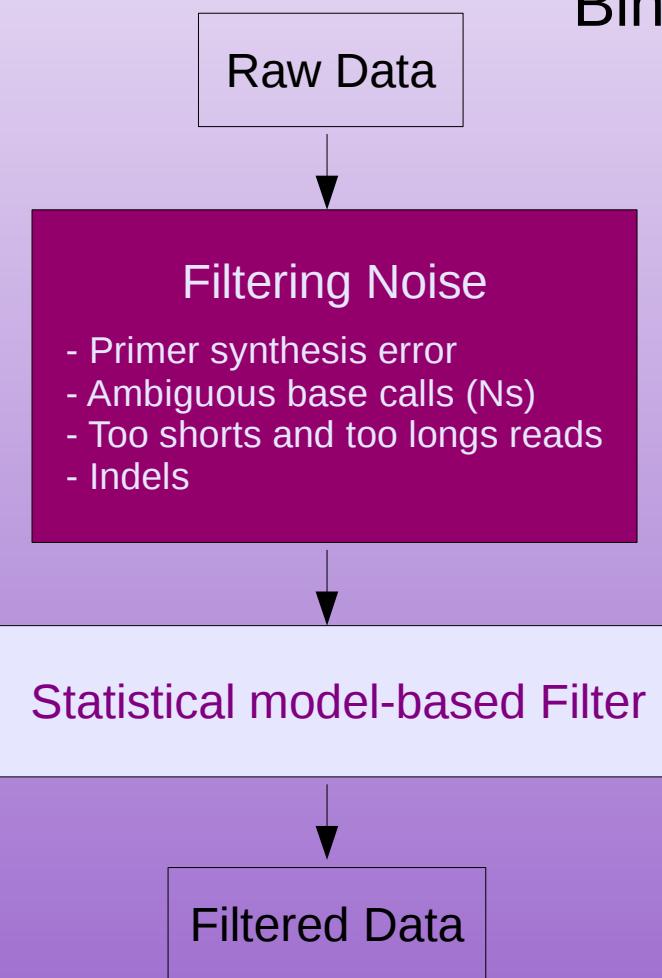
Filtering noise



- Remove reads with more than 25% of primer synthesis errors
 - Remove reads with ambiguous base calls (N)
 - Remove short reads and trim too long ones
 - Remove reads containing indels
- **(Huse et al. *Gen. Biol.* 2007)**

Modelling the remaining error

- Mismatch errors per site can be modeled with a Binomial distribution



... GGCAGGGGCAGCCTGCTTCGCCCGGCCATCTC ...
 ... GGCAGGGGCAGCCTGCTTCGCCCGGCCATCTC ...
 ... GGCAGGGGCAGCCTGCTTCGCCCGGCCATCTC ...
 ... GGCAGGGGCAGCCTGCTTCGCCCGGCCATCTC ...
 ... GGCAGGGGCAGCCTGCTATGCCCGGCCATCTC ...
 ... GGCAGGGGCAGCCTGCTTCGCCCGGCCATCTC ...

 ... GGCAGGGGCAGCCTGCTTCGCCCGGCCATCTC ...
 ... GGCAGGGGCAGCCTGCTATGCCCGGCCATCTC ...
 ... GGCAGGGGCAGCCTGCTTCGCCCGGCCATCTC ...

$\brace{N \text{ reads}}$

$$\text{Bin}(N,p) \approx \text{Poiss}(Np) \quad (N \rightarrow \infty, p \rightarrow 0)$$

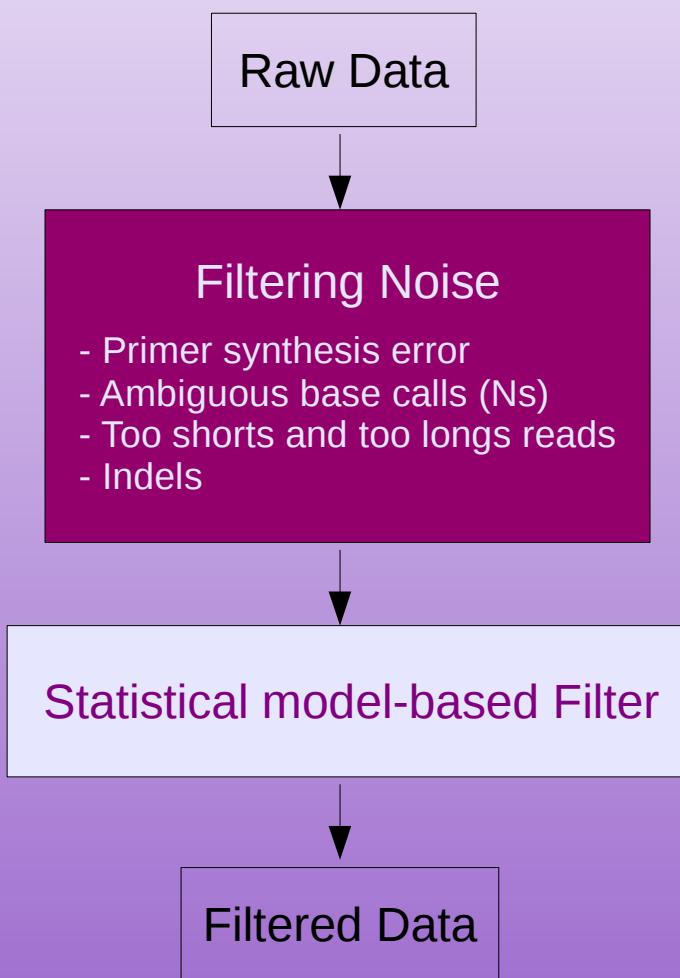
where $p = \text{prob}\{\text{mismatch}\}$,
 $N = \text{number of reads}$

Modelling the remaining error

- Modelling error: **(Wang et al. Gen. Res. 2007)**
- Mismatch errors $\sim \text{Poiss}(\lambda_r)$, $r \in \{\text{"h"}, \text{"nh"}\}$
- For a variant observed n times: the probability of getting it $\geq n$ times if it was an error is given by:

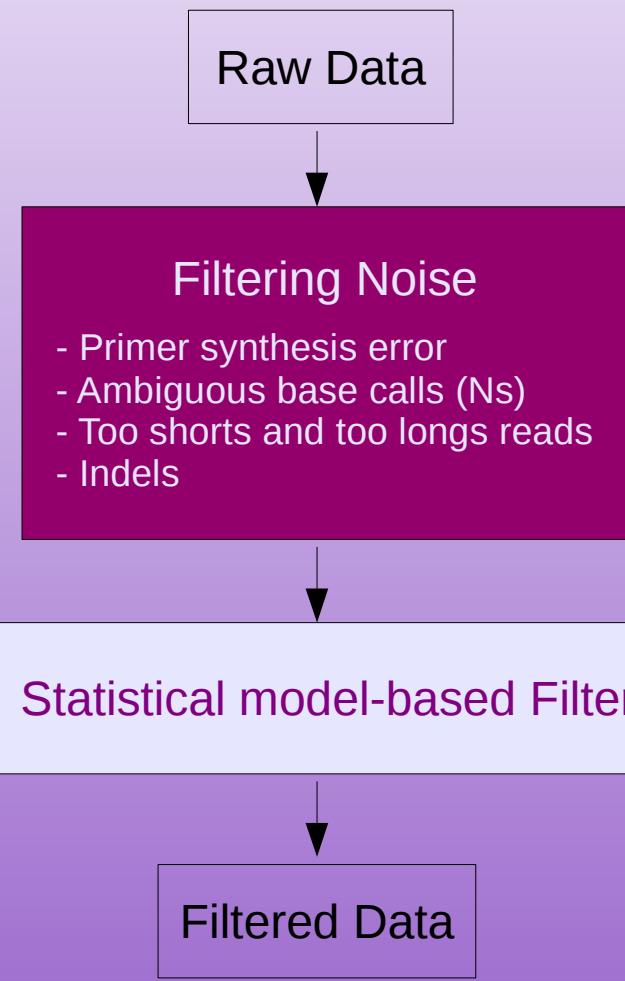
$$P = 1 - \sum_{i=1}^{n-1} \frac{e^{-\lambda} \cdot \lambda^i}{i!}$$

$$\lambda = N \cdot \lambda^r, r \in \{h, nh\}$$



Modelling the remaining error

- For a variant observed n times: the probability of getting it $\geq n$ times if it was an error is given by:



$$P = 1 - \sum_{i=1}^{n-1} \frac{e^{-\lambda} \cdot \lambda^i}{i!}$$

$$\lambda = N \cdot \lambda_{ij}^r, r \in \{h, nh\}, i, j \in \{A, C, T, G\}$$

- Matrices of mismatch counts and error rates:

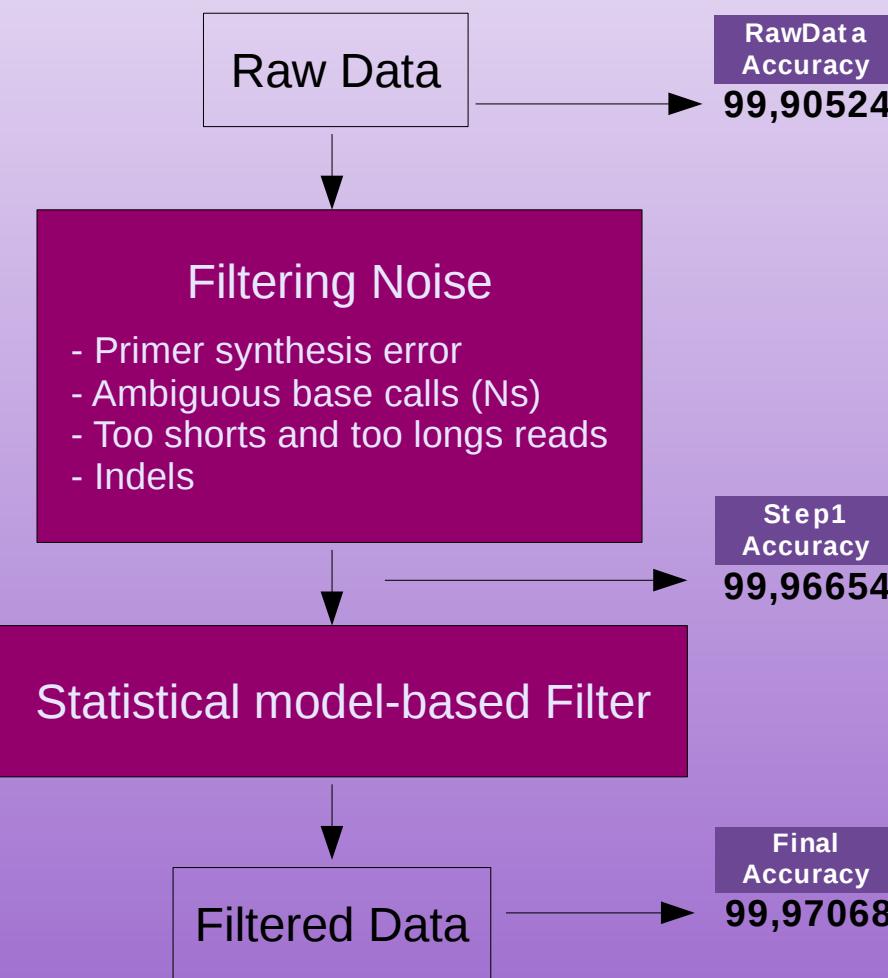
B12C1				
B12C1(H)				
	A	C	T	G
A	80714	2	7	7
C	19	209807	69	3
T	12	2	96858	4
G	136	3	6	161315

B12C1(NH)				
	A	C	T	G
A	113004	2	8	8
C	24	252892	36	2
T	6	9	166825	2
G	155	4	10	263549

B12C1				
B12C1 (H)				
	A	C	T	G
A	0,9998018	0,0000248	0,0000867	0,0000867
C	0,0000905	0,9995665	0,0003287	0,0000143
T	0,0001239	0,0000206	0,9998142	0,0000413
G	0,0008423	0,0000186	0,0000372	0,9991019

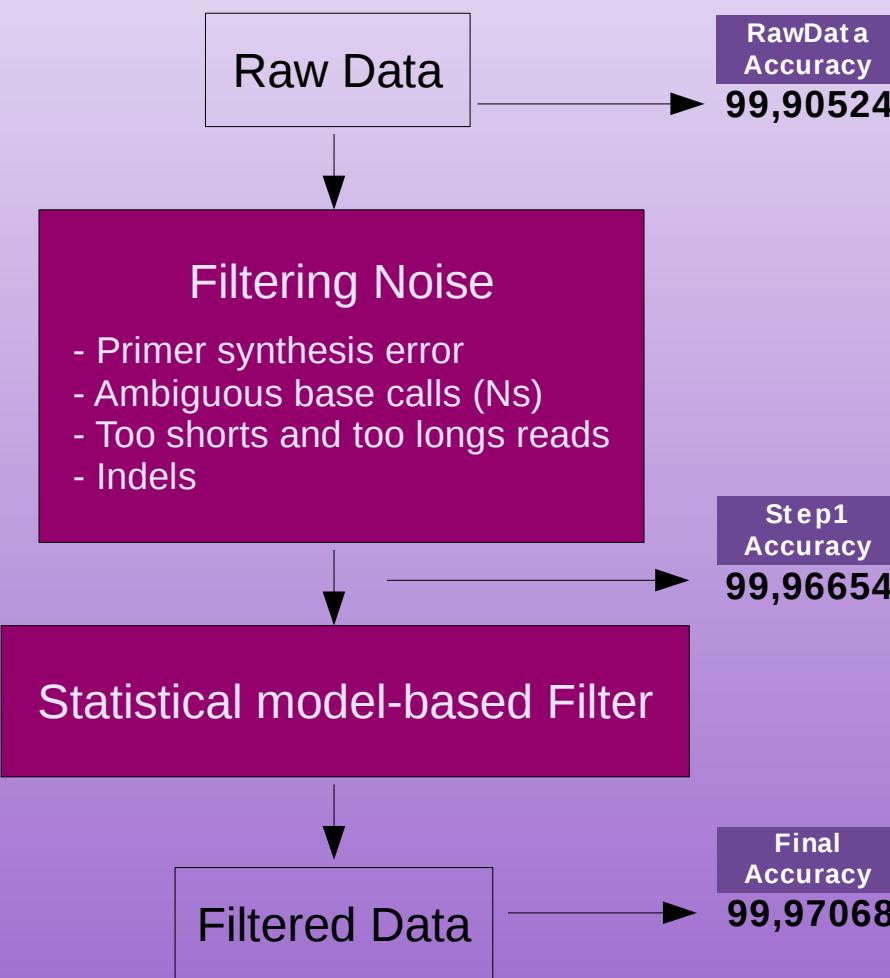
B12C1 (NH)				
	A	C	T	G
A	0,9998407	0,0000177	0,0000708	0,0000708
C	0,0000949	0,9997549	0,0001423	0,0000079
T	0,0000360	0,0000539	0,9998981	0,0000120
G	0,0005877	0,0000152	0,0000379	0,9993592

Results and conclusions



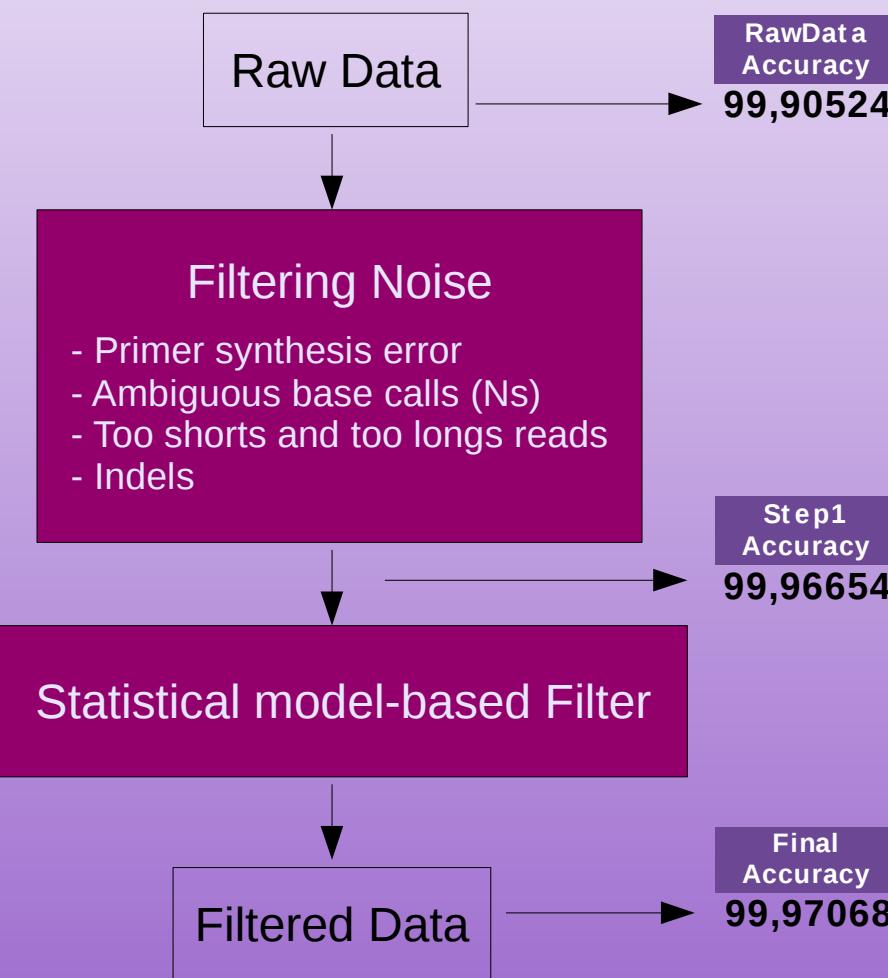
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Results and conclusions



- Filtering noise before computing estimations of proportions is a useful approach
- The statistical model:
 - improves slightly the accuracy
 - It provides a probabilistic score per base

Results and conclusions



- Filtering noise before computing estimations of proportions is a useful approach
- The statistical model:
 - improves slightly the accuracy
 - It provides a probabilistic score per base
- We still have to fine tune some points but we think that this pipeline could be useful in detecting minor variants.

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Thanks for your attention!